A Lead-Filled G-Quadruplex: Insight into the G-Quartet's Selectivity for Pb^{2+} over K^+

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ABSTRACT



The lipophilic nucleoside, G 1, extracts Pb^{2+} picrate from water into organic solvents to give structures based on the hydrogen-bonded G-quartet. Crystal structures indicate important differences between (G 1)₈-Pb²⁺ and (G 1)₈-K⁺. The divalent Pb²⁺ templates a smaller G₈ cage than does K⁺, as judged by the M–O6 bond length, O6–O6 diagonal distance, and inter-tetramer separation. The more compact Pb²⁺ octamer correlates with NMR data indicating that N2–N7 hydrogen bonds in (G 1)₈-Pb²⁺ are kinetically more stable than in (G 1)₈-K⁺.

With the increasing activity in supramolecular chemistry,¹ nucleobases have been used to construct some interesting and functional noncovalent assemblies.^{2–6} In addition to making new supramolecular architectures, lipophilic nucleobases also serve as valuable models for better understanding the factors that control structure and dynamics in duplex,⁷

- (3) Bell, T. W.; Hou, Z.; Zimmerman, S. C.; Thiessen, P. A. Angew. Chem., Int. Ed. Engl. 1995, 34, 2163-2165.
- (4) Schall, O. F.; Gokel, G. W. J. Am. Chem. Soc. 1994, 116, 6089-6100.
- (5) (a) Sigel, R. K. O.; Freisenger, E.; Metzger, S.; Lippert, B. J. Am. Chem. Soc. **1998**, *120*, 12000–12007. (b) Sigel, R. K. O.; Lippert, B. Chem. Commun. **1999**, 2167–2168.

(6) (a) Egholm, M.; Buchardt, O.; Nielsen, P. E.; Berg, R. H. J. Am. Chem. Soc. **1992**, 114, 1895–1897. (b) Egholm, M.; Nielsen, P. E.; Buchardt, O.; Berg, R. H. J. Am. Chem. Soc. **1992**, 114, 9677–9678.

(7) (a) Williams, N. G.; Williams, L. D.; Shaw, B. R. J. Am. Chem. Soc. **1989**, 111, 7205–7209. (b) Williams, L. D.; Williams, N. G.; Shaw, B. R. J. Am. Chem. Soc. **1989**, 112, 829–833. triplex,⁸ and tetraplex^{9,10} nucleic acids. Both Gottarelli's group and our group have been studying the cation-templated self-association of lipophilic guanosine derivatives.^{10–12} These compounds form self-assembled ionophores that bind cations with affinity and selectivity. In addition to represent-

⁽¹⁾ Reinhoudt, D. N.; Stoddart, J. F.; Ungaro, R. Chem. Eur. J. 1998, 4, 1349–1351.

^{(2) (}a) Sessler, J. L.; Wang, B.; Harriman, A. J. Am. Chem. Soc. **1995**, 117, 704–714. (b) Harriman, A.; Kubo, Y.; Sessler, J. L. J. Am. Chem. Soc. **1992**, 114, 388–390.

⁽⁸⁾ Zimmerman, S. C.; Schmitt, P. J. Am. Chem. Soc. 1995, 117, 10769–10770.

⁽⁹⁾ Sessler, J. L.; Sathiosatham, M.; Doerr, K.; Lynch, V.; Abboud, K. A. Angew. Chem., Int. Ed. Engl. **2000**, *39*, 1300–1303.

^{(10) (}a) Forman, S. L.; Fettinger, J. C.; Pieraccini, S.; Gottarelli, G.; Davis, J. T. J. Am. Chem. Soc. **2000**, 122, 4060–4067. (b) Marlow, A. L.; Mezzina, E.; Spada, G. P.; Masiero, S.; Davis, J. T.; Gottarelli, G. J. Org. Chem. **1999**, 64, 5116–5123.

^{(11) (}a) Gottarelli, G.; Masiero, S.; Spada, G. P. *Chem. Commun.* **1995**, 2555–2557. (b) Gottarelli, G.; Mariani, P.; Masiero, S.; Mezzina, E.; Spada, G. P.; Recanatini, M. *Helv. Chim. Acta* **1998**, *81*, 2078–2092. (c) Andrisano, V.; Gottarelli, G.; Masiero, S.; Heijne, E. H.; Pieraccini, S.; Spada, G. P. *Angew. Chem., Int. Ed.* **1999**, *38*, 2386–2388.

^{(12) (}a) Cai, M.; Marlow, A. L.; Fettinger, J. C.; Fabris, D.; Haverlock, T. J.; Moyer, B. A., Davis, J. T. Angew. Chem., Int. Ed. **2000**, 39, 1283– 1285. (b) Cai, M.; Sidorov, V.; Lam, Y. F.; Flowers, R. A.; Davis, J. T. Org. Lett. **2000**, 2, 1665–1668. (c) Davis, J. T.; Tirumala, S.; Marlow, A. L. J. Am. Chem. Soc. **1997**, 119, 5271–5272. (d) Davis, J. T.; Tirumala, S.; Jenssen, J. R.; Radler, E.; Fabris, D. J. Org. Chem. **1995**, 60, 4167– 4176.

ing a new approach toward ion coordination in organic solvents, our studies should also provide insight into the structural properties of higher-ordered nucleic acid aggregates. Herein, we report solid-state and solution evidence for complexation of Pb^{2+} by the lipophilic guanosine analogue G **1**. The resulting (G **1**)₈-Pb²⁺ octamer is a sandwich of two hydrogen-bonded G-quartets (Scheme 1).



The G-quartet is templated and stabilized by cations.¹³ In G-rich DNA, contiguous G-quartets stack to give structures known as G-quadruplexes.¹⁴ In addition to an affinity for the monovalent K⁺ and Na⁺, the G-quartet also binds divalent cations such as Ba²⁺ and Sr^{2+,15,16} Earlier this year, Smirnov and Shafer reported that Pb²⁺ is significantly better than K⁺ at inducing G-quartet structure in a DNA oligonucleotide.¹⁷ Since Pb²⁺ (r = 1.29 Å) has an ionic radius similar to but smaller than that of K⁺ (r = 1.51 Å),¹⁸ Pb²⁺ should fit into the G₈ cage formed by two stacked G-quartets. It is also reasonable that divalent metal ion coordination might further stabilize the G-quartet's hydrogen bonds. For example, calculations predict that cation coordination to G strengthens hydrogen bonds in G–C and G–G base pairs.¹⁹ This polarization enhancement of hydrogen bond strength is

calculated to be greater for divalent cations than for monovalent cations.²⁰ Such polarization effects have been considered when explaining the G-quartet's monovalent cation selectivity.²¹

In addition to providing general insight into G-quartet– cation interactions, studying Pb²⁺ binding to nucleobases may result in a better understanding of the molecular basis for lead's genotoxicity.²² While lead's coordination chemistry is well understood,²³ there are few structural details regarding binding of Pb²⁺ to nucleic acids. X-ray crystallography has shown that tRNA and RNA leadzymes have specific Pb²⁺ binding sites that utilize both nucleobase and phosphate ligands.^{24,25} Shafer's finding that Pb²⁺ promotes folding of a DNA G-quadruplex¹⁷ focused our attention on the interaction of Pb²⁺ with the lipophilic G **1**.

The nucleoside G **1** is an excellent model compound for obtaining molecular level details about G-quartets. We recently reported a G-quadruplex crystal structure formed from G **1** and K⁺ picrate.^{10a} The G-quadruplex was composed of two coaxial (G **1**)₈-K⁺ octamers with K⁺ cations sandwiched between G-quartet layers. We have now located Pb²⁺ cations within a similar G-quadruplex. Our current solid-state and solution data confirm that Pb²⁺ is better than K⁺ at stabilizing the G-quadruplex.

Solid State Structure. The lipophilic G **1** extracted Pb²⁺ picrate into CDCl₃ from water containing a 1:2 molar ratio of PbCl₂ and K⁺ picrate.²⁶ Integration of ¹H NMR signals for G **1** and picrate indicated an octameric stoichiometry. Solvent evaporation gave a solid whose elemental analysis was consistent with (G **1**)₈-Pb²⁺(pic)₂.²⁷ Single crystals, from CH₃CN/CHCl₃, had unit cell dimensions that were macromolecular: a = 25.5691(13) Å, b = 44.385(2) Å, and c = 83.840(4) Å.²⁸ This cell contained four G-quadruplexes, representing over 4500 non-hydrogen atoms. Each G-quadruplex was formed from two coaxial (G **1**)₈-Pb²⁺

(20) Spöner, J.; Burda, J. V.; Mejzlik, P.; Leszczynski, J.; Hobza, P. J. Biomol. Struct. Dyn. 1997, 14, 613–628.
(21) (a) Ross, W. S.; Hardin, C. C. J. Am. Chem. Soc. 1994, 116, 6070–

(21) (a) Ross, W. S.; Hardin, C. C. J. Am. Chem. Soc. **1994**, *116*, 6070–6080. (b) Tohl, J.; Eimer, W. Biophys. Chem. **1997**, *67*, 177–186. (c) Spackova, N.; Berger, I.; Spöner, J. J. Am. Chem. Soc. **1999**, *121*, 5519–

5534. (d) Gu, J.; Leszczynski, J. J. Phys. Chem. A 2000, 104, 6308-6313.
(22) For a review, see: Hartwig, A. Environ. Health Perspect. 1994, 102(S3), 45-50.

(23) Shimoni-Livny, L.; Glusker, J. P.; Bock, C. W. Inorg. Chem. 1998, 37, 1853–1867.

(24) Brown, R. S.; Hingerty, B. E.; Dewan, J. C.; Klug, A. *Nature* **1983**, *303*, 543–546.

(25) Wedekind, J. E.; McKay, D. B. Nat. Struct. Biol. 1999, 6, 261-268.

(26) Competitive extraction experiments indicate that G 1 has at least a 100:1 extraction selectivity for Pb^{+2} picrate over K^+ picrate.

(27) Calculated for $C_{164}H_{252}N_{46}O_{54}Si_8Pb: C, 47.32$; H, 6.06; N, 15.48; Pb, 4.98. Found: C, 47.19; H, 6.18; N, 15.42; Pb, 5.01.

(28) Crystal data for (G 1)₈·Pb(pic)₂·CH₃CN_{2.6}·[CHCl₃]_{3.6}·H₂O_{4.6}: C_{169.81}-H_{270.06}N_{48.63}O_{58.63}Cl_{1.31}Si₈Pb, MW = 4409.35, orthorhombic, space group P2₁₂1₂; a = 25.5691(13), b = 44.385(2), and c = 83.840(4) Å; $\alpha = 90^{\circ}$, $\beta = 90^{\circ}$, $\gamma = 90^{\circ}$; V = 95, 149(8) Å³; Z = 16, μ (Mo K_{α}) = 0.845 mm⁻¹. Data were collected at 193(2) K on a Bruker SMART1000 CCD diffractometer. The structure was determined by direct methods and refined using SHELXL.³⁶ The structure was refined to convergence with R(F) = 16.19%, wR(F²) = 24.35%, and GOF = 1.037 for 88,730 independent reflections [R(F) = 8.75%, wR(F²) = 21.40% for those 54223 data with $F_o > 4(F_o)$]. Crystal data (excluding structure factors) are deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC-146884. Data can be obtained free from the CCDC, 12 Union Road, Cambridge CB2 1EZ, U.K. (fax: (+44)1223-336-033. e-mail: deposit@ccd.cam.ac.uk).

⁽¹³⁾ For reviews, see: (a) Guschlbauer, W.; Chantot, J. F.; Thiele, D. J. Biomol. Struct. Dynam. **1990**, 8, 491–511. (b) Gilbert, D. E.; Feigon, J. Curr. Opin. Struct. Biol. **1999**, 9, 305–314.

⁽¹⁴⁾ For X-ray crystal structures of DNA G-quadruplexes, see: (a) Kang,
C.; Zhang, X.; Ratliff, R.; Moyzis, R.; Rich, A. Nature 1992, 356, 126–131. (b) Laughlan, G.; Murchie, A. I. H.; Norman, D. G.; Moore, M. H.; Moody, P. C. E.; Lilley, D. M. J.; Luisi, B. Science 1994, 265, 520–524. (c) Phillips, K.; Dauter, Z.; Murchie, A. I. H.; Lilley, D. M. J.; Luisi, B. J. Mol. Biology 1997, 273, 171–182.

⁽¹⁵⁾ Chantot, J.-F.; Guschlbauer, W. FEBS Lett. 1969, 4, 173-176.

⁽¹⁶⁾ Chen, F. M. Biochemistry 1992, 31, 3769-3776.

⁽¹⁷⁾ Smirnov, I.; Shafer, R. H. J. Mol. Biol. 2000, 296, 1-5.

⁽¹⁸⁾ These values are for the octa-coordinate cations: Shannon, R. D. *Acta Crystallogr. A* **1976**, *32*, 751–767.

⁽¹⁹⁾ Burda, J. V.; Spöner, J.; Leszczynski, J.; Hobza, P. J. Phys. Chem. B 1997, 101, 9670-9677.

octamers (Figure 1A). With individual G-quartets twisted 30° relative to each other, each Pb²⁺ cation coordinates eight



Figure 1. (A) Ball-and-stick representation of the lead-filled G-quadruplex. This G-quadruplex is composed of two coaxial (G 1)₈-Pb²⁺ octamers. The individual G-quartets, $G_4 \ 1-G_4 \ 4$, are labeled. The picrate anions are removed for clarity. (B) This space-filling representation of the octamer (G 1)₈-Pb²⁺ shows the eight oxygen atoms in the twisted G_8 cage coordinated to Pb²⁺. Average hydrogen bond distances, Pb²⁺-O6 distances, and G_4-G_4 inter-quartet distances for (G 1)₈-Pb²⁺ units are listed in Table 1.

O6 atoms in a geometry intermediate between a cube and a square antiprism (Figure 1B). Overall, the crystal structures for the $K^{+ 10a}$ and Pb^{2+} quadruplexes are quite similar, raising the issue of whether the genotoxicity of Pb^{2+} may be due to its ability to substitute for K^{+} in nucleic acid structures.

Despite their similarities, the $(G \ 1)_8$ -Pb²⁺ and $(G \ 1)_8$ -K⁺ units have some key structural differences consistent with Pb²⁺ forming the more stable octamer (Table 1). First, the

Table 1. Mean Distances (Å) in the (G 1)₈ Octamer Units from X-ray Crystal Structures of the Pb^{2+} and K^+ G-Quadruplexes^{*a*,*b*}

	(G 1) ₈ -Pb ²⁺	(G 1) ₈ -K ⁺
M-06	2.66 ± 0.05	2.80 ± 0.06
O6-O6	4.46 ± 0.05	4.58 ± 0.06
between (G 1) ₄ planes	3.22 ± 0.01	3.31 ± 0.03
N1-O6 H-bond	2.86 ± 0.03	2.88 ± 0.02
N2–N7 H-bond	2.82 ± 0.02	2.90 ± 0.01

^{*a*} Values for (G 1)₈-K⁺ are mean distances for the unit cell's four G-quartets, see ref 10a. The standard deviations are those observed for the set of distances in the four G-quartets. ^{*b*} Values for (G 1)₈-Pb²⁺ are mean distances for the structure's 16 unique G-quartets. The standard deviations are those observed for the set of distances in the 16 G-quartets.

mean cation—G O6 distances are 0.14 Å shorter in (G 1)₈-Pb²⁺ than in (G 1)₈-K⁺. Second, the mean O6—O6 diagonal, a measure of G-quartet diameter,²⁹ is 0.12 Å shorter for (G 1)₈-Pb²⁺ than for (G 1)₈-K⁺. Third, vertical separation of G-quartets in (G 1)₈-Pb²⁺ is approximately 0.10 Å less than in (G 1)₈-K⁺. In short, the divalent Pb²⁺ templates a smaller G₈ cage than does K⁺. Hydrogen bond lengths for the N2

 H_A -N7 pair also become shorter as the octamer cage shrinks. As described below, a more compact octamer correlates well with NMR data indicating that the N2 H_A -N7 hydrogen bonds in (G 1)₈-Pb²⁺ are kinetically more stable than in (G 1)₈-K⁺.

Solution NMR Studies. We used both ²⁰⁷Pb and ¹H NMR to show that the (G 1)₈-Pb²⁺ is also stable in solution. Previous heteronuclear NMR studies using ²³Na⁺, ¹⁵NH₄⁺, and ⁸¹Tl⁺ have directly demonstrated cation binding by DNA G-quartets.^{30–32} Lead-207, a spin ¹/₂ nucleus of 22% natural abundance, has a large chemical shift range (16 000 ppm) that makes its NMR spectra exquisitely sensitive to the coordination environment.³³ After extraction of Pb²⁺ picrate by G 1, a sharp ²⁰⁷Pb NMR signal in CDCl₃ was observed at δ –3029, relative to PbMe₄ (see Supporting Information). The same ²⁰⁷Pb NMR peak was observed when crystals of the Pb²⁺ complex were dissolved in CDCl₃. This ²⁰⁷Pb NMR peak is strong evidence for cation coordination by G 1, since Pb²⁺ picrate itself is insoluble in CDCl₃.

Two sets of ¹H NMR signals in a 1:1 ratio and diagnostic NOEs revealed that (G 1)₈-Pb²⁺ forms in CDCl₃ by head-to-tail stacking of G-quartets.³⁴ Amide N1 H (δ 11.80 and 11.41) and amino N2 H_A (δ 9.97 and 9.20) resonances were downfield shifted, as expected for hydrogen-bonded protons. These resonances were present only after Pb²⁺ extraction, again strong evidence that the cation templates the G-quartet's structure.

In the ¹H NMR spectrum of a G₈ octamer, there are two sets of amino resonances. Each set contains a hydrogenbonded resonance (N2 H_A) and a non-hydrogen-bonded resonance (N2 H_B). The ¹H NMR spectra revealed that Pb²⁺, as compared to K⁺, forms a G-quartet with kinetically stronger N2 H_A-N7 hydrogen bonds. Specifically, C2–N2 bond rotation was slower in (G 1)₈-Pb²⁺ than in (G 1)₈-K⁺.

All four amino NH₂ resonances in (G 1)₈-Pb²⁺ were sharp and distinct at 25 °C (Figure 2). Coalescence of these amino signals did not occur even at 50 °C, indicating a significant barrier for C2–N2 bond rotation in (G 1)₈-Pb²⁺. In marked contrast, amino resonances for (G 1)₈-K⁺ were broadened into the baseline at temperatures above 10 °C, indicating



Figure 2. A region of the 500 MHz ¹H NMR spectra of (G 1)₈-Pb²⁺(pic)₂ (5 mM) in CDCl₃ at 25 °C. The two sets of separate resonances for the N2 H_A and N2 H_B amino protons (marked by asterisks) indicate a significant barrier for C2–N2 bond rotation in (G 1)₈-Pb²⁺.

⁽²⁹⁾ Strahan, G. D.; Keniry, M. A.; Shafer, R. H. Biophys. J. 1998, 75, 968–981.

much faster C2–N2 bond rotation in the K⁺ octamer. A conservative estimate indicates that ΔG_c^{\dagger} for C–N bond rotation is at least 1.5 kcal/mol greater for the Pb²⁺ complex as compared to the K⁺ complex.³⁵ These results, showing that the C–N bond rotation barrier is significantly higher for (G 1)₈-Pb²⁺ relative to (G 1)₈-K⁺, indicate that the divalent cation stabilizes the G-quartet's hydrogen bonds more than a monovalent cation.

Conclusion. Both the solid state and solution evidence show that the smaller and more highly charged Pb^{2+} cation templates a smaller G_8 octamer cage than does K^+ . This

(35) This estimate was made by assuming that the N2 H_A-H_B coalescence temperature is 50 °C for the Pb²⁺ complex (an underestimate) and 10 °C for the K⁺ complex. The equations $k_c = \pi \Delta v / \sqrt{2}$ and $\Delta G^{\dagger}_c = 2.3 R T_c$ -[10.32 + log T_c/k_c] were used to approximate C2–N2 rotation barriers of $\Delta G^{\dagger}_c = 13.5$ kcal/mol for the Pb²⁺ complex and $\Delta G^{\dagger}_c = 12.0$ kcal/mol for the K⁺ complex.

(36) Sheldrick, G. M. SHELXL-93 Program for the Refinement of Crystal Structures; University of Göttingen: Germany, 1993.

tighter coordination geometry kinetically stabilizes the Gquartet's N2 H_A –N7 hydrogen bonds. These experimental results, including data from the first crystal structure of a G-quadruplex bound to a divalent cation, are consistent with calculations that predict the polarization enhancement of DNA base pairing upon cation binding.^{19–21} While it remains to be seen if Pb²⁺ binding to DNA G-quartets has a role in the cause and effect of lead's genotoxicity, these studies with G **1** provide a firm rationale for why Pb²⁺ binds more tightly to a G-quadruplex than does K⁺.

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Supporting Information Available: Crystallographic tables, final coordinates and thermal parameters, selected bond lengths and angles, and ¹H and ²⁰⁷Pb NMR spectra. This material is available free of charge via the Internet at http://pubs.acs.org.

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⁽³⁰⁾ Deng, H.; Braunlin, W. H. J. Mol. Biol. **1996**, 255, 476–483.

⁽³¹⁾ Hud, N. V.; Schultze, P.; Feigon, J. J. Am. Chem. Soc. 1998, 120, 6403-6404.

⁽³²⁾ Basu, S.; Szewczak, A. A.; Cocco, M.; Strobel, S. A. J. Am. Chem. Soc. 2000, 122, 3240–3241.

⁽³³⁾ Claudio, E. S.; ter Horst, M. A.; Forde, C. E.; Stern, C. L.; Zart, M. K.; Godwin, H. A. *Inorg. Chem.* **2000**, *39*, 1391–1397.

⁽³⁴⁾ For a detailed NMR study of a "head-to-tail" dG_8 -K⁺ octamer from another lipophilic nucleoside, see ref 10b.